

Inflammatory, Apoptotic, and Survival Gene Signaling in Alzheimer's Disease

A Review on the Bioactivity of Neuroprotectin D1 and Apoptosis

Walter J. Lukiw · Nicolas G. Bazan

Received: 5 April 2010 / Accepted: 5 April 2010 / Published online: 23 April 2010
© Springer Science+Business Media, LLC 2010

Abstract Aging is associated with an enhanced susceptibility to brain dysfunction, loss of memory, and cognitive decline and significantly influences the quality of life for the affected individual. Recent molecular–genetic approaches have provided powerful insights into common age-related diseases that are both progressive and multifactorial, such as Alzheimer's disease (AD), and in vitro in AD models. These investigations have uncovered consistent deficits in brain gene signaling mechanisms and neurotrophic substances known to contribute to normal brain function. Inflammatory signaling pathways involving up-regulation of cytosolic phospholipase A₂ and the arachidonic acid cycle, the depletion of the brain-essential fatty acid docosahexaenoic acid (DHA) and DHA-derived neuroprotectin D1, and changes in the expression of key proapoptotic and antiapoptotic members of the Bcl-2 gene family are thought to be major contributors to pathogenic processes in degenerating brain tissue. This review will focus on the roles of stress genes, apoptosis-related genes, and inflammation in the molecular genetics of AD with emphasis on the interactive nature of inflammatory, neurotrophic, and apoptotic signaling and will highlight areas of rapid progress in the characterization of action of DHA and neuroprotectin D1 and address important research challenges. We also attempt to integrate these molecular, genetic, and neurochemical changes with cellular pathways

involved in brain aging to formulate an integrated understanding of multifactorial age-related neurologic disease and pharmacotherapeutic strategies that may be useful in the restoration of homeostatic brain function.

Keyword Alzheimer's disease (AD) · Apolipoprotein E4 (ApoE4) · Apoptosis · Bcl-2, docosahexaenoic acid (DHA) · Multifactorial · Neuroprotectin D1 (NPD1) · Phospholipase A₂ · Reactive oxygen species (ROS)

List of Abbreviations

Aβ ₄₂	amyloid beta 42-amino-acid peptide
AA	arachidonic acid
AD	Alzheimer's disease
ApoE4	apolipoprotein E4 allele
βAPP	beta-amyloid precursor protein
COX-2	inducible cyclooxygenase-2
cPLA ₂	cytosolic phospholipase A ₂
DHA	docosahexaenoic acid
HNE	hydroxynonenal
LOX	lipoxygenase
LPX	lipoxins
LTR	leukotrienes
NFT	neurofibrillary tangles
NPD1	neuroprotectin D1
PG	prostaglandin
PLA ₂	phospholipase A ₂
PUFA	polyunsaturated fatty acid
ROS	reactive oxygen species
SP	senile plaque
sAPPα	soluble amyloid precursor protein alpha fragment
sPLA ₂	secreted phospholipase A ₂
TNFAIP2	tumor necrosis factor alpha inducible protein-2 (B94)

W. J. Lukiw (✉) · N. G. Bazan
LSU Neuroscience Center of Excellence and Department
of Ophthalmology, Louisiana State University Health
Sciences Center,
2020 Gravier Street, Suite 904,
New Orleans, LA 70112-2272, USA
e-mail: wlukiw@lsuhsc.edu

N. G. Bazan
e-mail: nbazan@lsuhsc.edu

Introduction

Neuropathological and cognitive changes that accompany dementia syndromes in aged humans are progressive, interrelated, and highly complex. Alzheimer's disease (AD) represents the most prevalent neurologic dysfunction in aging Western and Asian populations, currently affecting about 6 million Americans, or about 2% to 3% of the populations of modern industrialized societies. Although the first case of AD was described over 103 years ago, the pathogenic evolution of AD is still not completely understood. Contributory and highly interrelated factors for AD include familial and inherited gene polymorphisms, environmental toxins, prior viral infection, cerebral hypoxia, ischemia and hypoperfusion, cerebrovascular and neurovascular disease, dysregulated proinflammatory signaling, and the appearance of molecular lesions that include deposition and aggregation of amyloid beta ($A\beta$) peptides as senile plaques (SPs) and tau-protein containing neurofibrillary tangles (NFTs; 1-5). Importantly, SP and NFT lesions show nonrandom patterns of deposition in the association neocortex and hippocampus, relatively recently evolved brain compartments in humans. These limbic regions are involved in higher cognitive function and memory and where AD-type neuropathological changes make their earliest as well as most severe appearance [1, 2]. Similarly, large pyramidal neurons in layers 3 and 5 of the association neocortex seem to be preferentially atrophied by the AD process and display significant loss of arborization [1–3]. Why these large pyramidal neurons are preferentially targeted by the AD process is not well understood, but may be related to their large relative surface area and high rates of metabolism and DNA transcription [4–8]. Aged and AD-affected brains both exhibit somewhat similar physiological and pathological changes; however, the extent and severity of these changes are greatly amplified in transition from normal, healthy aging to AD. Full genome-wide gene expression profiling studies using high-density DNA array technologies have indicated a generalized depression in the expression of brain-specific genetic information in AD when compared with cognitively normal, age-matched brain [5–12]. These changes include deficits in the abundance of RNAs encoding neurotrophic factors, synaptosomal and signaling elements, caspases and other apoptosis-related factors, metal ion regulatory proteins, and signaling pathways involved in energy metabolism [4–9]. Importantly, in AD, there appear to be specific defects in the mechanisms that regulate gene transcription, translation, and protein processing for structural molecules that define the cytoarchitecture, and thereby the synaptic circuitry, connectivity, and signaling capabilities of the

neural network. Cumulatively, these genetic deficiencies correlate well with the characteristic atrophy of vulnerable neurons and synaptic structures within AD-affected brain areas at postmortem examination and the degree of cognitive decline observed in premortem clinical testing [5, 6, 10]. In contrast to the approximate two thirds of the total expressed brain genome found to be repressed in AD, there exists an additional family of NF-kappaB up-regulated genes that encode interactive elements involved in stress response, proinflammatory, and proapoptotic signaling [4–6, 11–13].

Recent research evidence suggests that at least three major molecular–genetic pathways contribute to the progressively dysfunctional, gene-mediated pathogenesis observed in AD-affected brain: [14] inflammatory signaling pathways that involve an up-regulation of cytosolic phospholipase A_2 (cPLA₂) and the arachidonic acid (AA) cycle, [15] the depletion in critical brain regions of the essential fatty acid docosahexaenoic acid (DHA) and the 15-lipoxygenase (15-LOX) DHA-derived neuroprotectin D1 (NPD1), and [16] changes in the expression of key proapoptotic and antiapoptotic members of the ~25 member Bcl-2 gene family that regulate brain cell fate decisions and caspase-3-mediated events that trigger cytochrome *c* release and apoptosis (Fig. 1). Interestingly, caspases may have a proximal role in the progression of neurodegenerative disease and not just in the terminal stages of neurodegeneration. For example, increased expression and activation of caspases is observed in subjects with mild AD [17–20]. $A\beta$ peptide-triggered oxidative stress induces caspase activation through the intrinsic mitochondrial pathway, and activation of executioner caspases, particularly caspase-3, may preferentially promote amyloidogenic β APP cleavage, potentially resulting in a feed-forward cycle of $A\beta$ peptide production and caspase-3 activation [18, 19]. Tau is also a substrate for caspase-3, and cleavage of tau may promote $A\beta$ peptide aggregation, paired helical filament formation, and nerve terminal degeneration [18–20]. The benefit of inhibiting the intrinsic apoptotic biochemical cascade has been further demonstrated in a triple transgenic AD mouse model wherein overexpression of the antiapoptotic Bcl-2 gene blocked activation of caspase-9 and caspase-3; tau and β APP processing were suppressed, numbers of NFTs and $A\beta$ 42 peptide deposition were reduced, and memory performance was enhanced [19, 20]. As will be emphasized in the following review, these apoptotic signals and gene-mediated pathways are highly interwoven in the fabric of the AD process and represent several attractive pharmacotherapeutic targets for the clinical management of this devastating neurologic disorder.

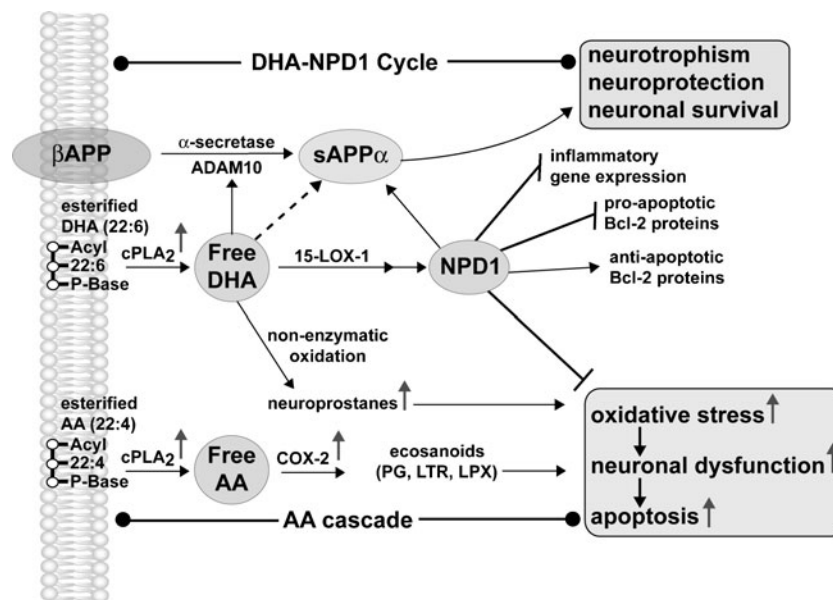


Fig. 1 Essentials of membrane-esterified DHA–free DHA and 15-LOX-mediated NPD1 generation and the AA cascade, including enzymatic and nonenzymatic processing pathways for free DHA. Liberation of free DHA and AA from phospholipid membrane stores is rate-limited by a tightly regulated brain-enriched, cPLA₂. DHA from the omega-3 essential fatty acid family preferentially accumulates within neuronal and retinal phospholipids of central nervous system membranes, concentrating specifically within neuronal plasma membranes, synapses, and retinal photoreceptors. Free DHA derived from membrane DHA stores (*upper*) liberated via cPLA₂ cleavage may subsequently be converted into the 10,17S docosatriene NPD1 through an enzyme-mediated lipoxygenation via a 15-LOX, or 15-LOX-like enzymes. Deficits in 15-LOX abundance correlate with low NPD1 levels in stressed primary brain cells and in AD brain tissue. The neurobiological activity of DHA-derived NPD1 in cultured human retinal pigment epithelial and human neural cells has been characterized as a potent cytoprotective and neuroprotective oxygenated lipid mediator. Some of these neurotrophic and neuroprotective properties may be mediated through an integral membrane protein β APP-derived sAPP α via a nonamyloidogenic, α -secretase (ADAM10)-mediated pathway that supports neurotrophism, neuroprotection, and neuronal survival. NPD1 may also directly stimulate the expression and activity of sAPP α . Neurotrophic support mediated by DHA and NPD1 enables homeostatic brain functions, and a DHA-

NPD1 cycle between exogenous DHA supplies and membrane stores is maintained. Membrane-esterified and free DHA is also rapidly oxidized nonenzymatically by molecular oxygen (O₂) and free nitrogen radicals to form F4 neuroprostanes, a class of peroxidized lipids that further support oxidative stress and cellular apoptosis leading to brain cell dysfunction. These nonenzymatic reactions may be quenched by specific antioxidants or free radical scavengers. Increases in cPLA₂ expression are highly characteristic of AD neuropathology. Free AA derived from membrane phospholipid stores (*lower*) liberated via cPLA₂ may be subsequently acted upon by a constitutive COX-1 or an inducible COX-2 enzyme to yield eicosanoids, including prostaglandins (PG), leukotrienes (LTR), lipoxins (LPX), and other highly active lipid mediators. When over-produced, these trigger oxidative stress, inflammatory signaling, and brain cell apoptosis leading to cellular dysfunction and apoptotic brain cell death. The specific paracrine interactions between neurons, astroglia, and endothelial cells of the neurovascular unit are just beginning to become understood [53]. Pharmacologic strategies that increase DHA and NPD1 signaling and quench AA-mediated pathogenic signaling might be expected to be beneficial to brain cells in reducing oxidative stress, brain cell dysfunction and apoptosis, and extend homeostatic neural network functions of the brain that include memory and cognition [54]. Some outstanding research questions are summarized in Table 1

Cytosolic Phospholipase A₂ and the Arachidonic Acid Cycle

A consistent observation in the neocortex and hippocampal region of AD affected brains is increases in the expression and activity of cytosolic phospholipase A₂ (cPLA₂), a ~100 kDa, intracellular, calcium-dependent membrane-associated esterase. As a brain-enriched member of a larger phospholipase A₂ gene family, cPLA₂ hydrolyzes the *sn*-2 position of membrane glycerophospholipids, liberating free arachidonic acid (AA), docosahexaenoic acid (DHA), and an

array of bioactive lipids that regulate vital aspects of neural membrane biology, including protein–lipid interactions, transmembrane, and transsynaptic signaling. An omega-6 fatty acid (20:4,n-6), AA is one of the most abundant lipids in the brain and is present in similar quantities to DHA (22:6,n-3), and these two polyunsaturated fatty acids account for approximately 20% of the brain's total lipid content [21]. Like DHA, which is discussed further below, neurologic health is highly reliant on sufficient levels of AA, which maintain hippocampal cell membrane fluidity and other brain essential signaling functions [21, 22].

Table 1 Interactions between the AA cascade, DHA, and NPD1 and other aspects of DHA-NPD1 metabolism in AD are not well understood, and several key questions remain to be answered (see also Fig. 1)

Outstanding Research Questions

- Does an impairment in NPD1 biosynthesis take place during the early development of AD?
 - Does NPD1 act at the convertase level to regulate ADAM10 maturation?
 - Does NPD1 modulate chaperone transcription related to β APP processing?
 - Is NPD1-mediated alpha secretase activation the driving mechanism of neurotrophism?
 - Are astrocytes, glial, or endothelial cells engaged in NPD1-mediated actions?
 - What mechanisms regulate cPLA₂ activity to specifically liberate DHA and AA?
 - Is the redox status of the cell an important regulating cPLA₂ activity or the presence of free, nondegraded DHA pools?
 - What signaling elements of the DHA-NPD1 cycle and the AA cascade are biologically interconnected?
-

Metabolites of cPLA₂ may act as second messengers themselves, or are further metabolized to yield bioactive lipid mediators such as free fatty acids, lysophospholipids, platelet-activating factor, eicosanoids such as the prostaglandins, and reactive oxygen species (ROS) from AA, the end-product of cPLA₂ action on membrane phospholipids, and the rate-limiting enzyme in the AA cycle. Over-activation of cPLA₂ is believed to be an important instigator of inflammatory brain diseases—interestingly, secretory PLA₂ (sPLA₂) are primary constituents of insect and snake venom that releases large amounts of AA from the phospholipid membrane stores at the site of injury resulting in a highly localized inflammatory response [23, 24]. Cyclooxygenase-2 (COX-2) and cPLA₂ are inducible brain enzymes that act in tandem on the breakdown of membrane glycerophospholipid stores interrelated bioactive lipids that promote neural inflammation and apoptotic signaling up-regulation in AD brain [25–31].

Docosahexaenoic Acid and Neuroprotectin D1 in Aging and in Alzheimer's Disease

The essential marine-derived omega-3 polyunsaturated fatty acid docosahexaenoic acid (DHA; 22:6n-3) is selectively concentrated in neuronal, synaptic, and retinal membranes. In fact up to 60% of fatty acids esterified in neuronal cell membrane phospholipid stores consist of DHA, so brain cells have a convenient and readily accessible supply of DHA, that through phospholipase activities, liberate DHA from membranes to serve in neural signaling, cell survival, and cell fate pathways. Stereospecific oxygenated derivatives of DHA created through 15-lipoxygenase (15-LOX) action on free DHA, further generate neuroprotectin D1 (NPD1) that elicits potent cytoplasmic, neural, and retinal protective effects. The neurophysiological actions of esterified DHA occur in part through the highly flexible nature of this 22:6n-3 fatty acid, the maintenance of plasma membrane integrity, and lipid bilayer biophysical effects,

improving neurotransmission via increased receptor binding and enhancement in the function of ion channels and affinity of receptors [32–34].

The beneficial actions of free DHA and NPD1 appear to mainly occur (a) through the repression of the induction of inflammatory signaling mediators such as the inducible COX 2 enzyme, (b) through the recruitment of antiapoptotic members of the Bcl-2 gene family, and (c) through the repression of proinflammatory and proapoptotic signaling genes and their translation products [34, 35]. AD exhibits a progressive deposition of ragged amyloid beta (A β) peptides derived from the beta-gamma (β - γ) secretase pathway that processes beta-amyloid precursor protein (β APP) into the more toxic forms of β APP-derived fragments. A β peptides themselves and downstream consequences of A β peptide signaling are pro-oxidative, neurotoxic, proinflammatory, and proapoptotic. The enzymatic generation, speciation, and trafficking of β APP and A β peptides in AD and in experimental AD models are impacted by DHA concentration, by the bioavailability of unesterified DHA, and by derivatives of DHA such as NPD1.

DHA is essential for prenatal brain and retinal development and normal, homeostatic cognitive, and visual functions. Dietary deficiencies in DHA are associated with neurologic and retinal dysfunction and cognitive and visual decline. Deficits in DHA and NPD1 abundance are associated with the neurodegenerative mechanisms that characterize AD [35–38]. DHA levels are lower in the blood plasma and brains of AD patients, which may be the result from low dietary intake and/or enhanced oxidation of these highly labile polyunsaturated fatty acids [35, 38]. Moreover, diets enriched in DHA reduces amyloid burden in an aged AD transgenic mouse models [32–34]. Interestingly, apolipoprotein E4 (apoE) genotype influences responsiveness to dietary DHA treatment and DHA supplementation to patients containing ApoE4 genotypes are not as effective as patients with “lower risk” genotypes, such as those homozygous for ApoE3 [39–42]. In a more

recent study of 2,233 at-risk patients in four different US communities, the consumption of fatty fish was associated with a reduced risk of dementia and AD for those without the ApoE4 allele; carriage of an ApoE4 allele greatly reduced or eliminated DHA benefit in these patients [40]. A recent review that includes the results of several additional DHA clinical trials has recently appeared in the literature [41].

NPD1 and Activation of Antiapoptotic Members of the Bcl-2 Gene Family

Neurons cease to function and die via necrotic or apoptotic mechanisms, depending on the nature and severity of the physiological insult. In contrast to necrosis, a form of traumatic cell death that results from acute ischemic or traumatic brain injury, apoptosis is a process of programmed cell death that involves a series of biochemical events that lead to a variety of morphologic changes, including cellular blebbing, loss of membrane asymmetry and attachment, cell shrinkage, nuclear fragmentation, chromatin condensation, and chromosomal DNA fragmentation. However, the boundaries between these cell death mechanisms are not always distinct, and crosstalk between necrosis and apoptosis is apparent [42–44].

In aging and the more chronic and progressive AD, neuronal cell death by apoptosis is thought to be the prominent mechanism of brain cell functional decline and death. The morphologic and biochemical features and mechanisms of the different phases of apoptotic neuron death are overlapping and progressive until neuronal death occurs [44, 45]. Key proapoptotic and antiapoptotic members of the ~25-member Bcl-2 gene family regulate brain cell fate decisions, and various caspase-3-mediated events trigger cytochrome *c* release and apoptotic events. The Bcl-2 gene family can be roughly separated into two groups—those that are proapoptotic, including the members Bax, BAD, Bak and Bok, and those that are antiapoptotic, including the members Bcl-2, Bcl-xL, Bcl-w, Bfl-1(A1), and others. Bcl-2 family members appear to govern mitochondrial membrane permeabilization, achieved in part by activation or inactivation of inner mitochondrial permeability transition channels involved in the regulation of mitochondrial matrix pH and calcium levels. Bcl-2 family proteins induce (proapoptotic members) or inhibit (antiapoptotic members) the release of mitochondrial bound cytochrome *c* into the cytosol where it activates caspase-3 and caspase-9 that leads to apoptosis and cellular demise [46–48]. Interestingly, NPD1 has been shown to both up-regulate the antiapoptotic genes encoding Bcl-2, Bcl-xL, and Bfl-1(A1) in human brain cells in culture and to down-regulate the proapoptotic genes Bax and Bik [48–52]. In parallel with these beneficial effects on Bcl-2 family gene expression, NPD1 also down-regulated the expression of

interleukin-1beta (IL-1 β), tumor necrosis factor alpha (TNF α), cyclooxygenase-2 (COX-2), and the tumor necrosis factor alpha inducible protein-2 B94 (TNFAIP2), all members of a proinflammatory gene family known to be up-regulated in AD brain [4–9].

Summary

Due to the high metabolic index of nervous tissues, brain cells and systems require sustained antioxidative and neurotrophic support to maintain their homeostatic signaling functions. Neurodegenerative disorders such as Alzheimer's disease (AD) are associated with dysfunction and decline of key, highly interconnected pyramidal cell groups in the association neocortex and hippocampus, in part through the appearance of insoluble SP and NFT deposits, and the proinflammatory and proapoptotic pathology that these insoluble pathogenic lesions induce. Administration of DHA in clinical trials and NPD1 in cellular studies has shown multifaceted benefit in the preservation of cognition, memory, and healthy brain cell function. Lifelong impairment in the supply of DHA and NPD1 to the brain might be expected to result in a chronic loss of neurotrophic support for nervous tissues and contribute to progressive disturbances in cognition, memory, and associated higher brain functions.

References

1. Lukiw WJ, Bazan NG (2008) Docosahexaenoic acid and the aging brain. *J Nutr* 138:2510–2514
2. Mann DM (1996) Pyramidal nerve cell loss in Alzheimer's disease. *Neurodegeneration* 5:423–427
3. Thangavel R, Sahu SK, Van Hoesen GW, Zaheer A (2009) Loss of nonphosphorylated neurofilament immunoreactivity in temporal cortical areas in Alzheimer's disease. *Neuroscience* 160:427–433
4. Colangelo V, Schurr J, Ball MJ, Pelaez RP, Bazan NG, Lukiw WJ (2002) Gene expression profiling of 12633 genes in Alzheimer hippocampal CA1: transcription and neurotrophic factor down-regulation and up-regulation of apoptotic and pro-inflammatory signaling. *J Neurosci Res* 70:462–473
5. Cui JG, Hill JM, Zhao Y, Lukiw WJ (2007) Expression of inflammatory genes in the primary visual cortex of late-stage Alzheimer's disease. *NeuroReport* 18:115–119
6. Webster JA, Gibbs JR, Clarke J, Ray M, Zhang W, Holmans P, Rohrer K, Zhao A, Marlowe L, Kaleem M, McCorquodale DS 3rd, Cuello C, Leung D, Bryden L, Nath P, Zismann VL, Joshipura K, Huentelman MJ, Hu-Lince D, Coon KD, Craig DW, Pearson JV, NACC-Neuropathology Group, Heward CB, Reiman EM, Stephan D, Hardy J, Myers AJ (2009) Genetic control of human brain transcript expression in Alzheimer disease. *Am J Hum Genet* 84:445–458
7. Liang WS, Duncley T, Beach TG, Grover A, Mastroeni D, Walker DG, Caselli RJ, Kukull WA, McKeel D, Morris JC, Hulette C, Schmechel D, Alexander GE, Reiman EM, Rogers J, Stephan DA (2006) Gene expression profiles in anatomically and functionally distinct regions of the normal aged human brain. *Physiol Genomics* 28:311–322

8. Lukiw WJ (2004) Gene expression profiling in fetal, aged, and Alzheimer hippocampus: a continuum of stress-related signaling. *Neurochem Res* 29:1287–1297
9. Boetkjaer A, Boedker M, Cui JG, Zhao Y, Lukiw WJ (2007) Synergism in the repression of COX-2- and TNF α -induction in platelet activating factor-stressed human neural cells. *Neurosci Lett* 426:59–63
10. Lukiw WJ, Rogaev EI, Bazan NG (1996) Synaptic and cytoskeletal RNA message levels in sporadic Alzheimer neocortex. *Alzheimer's Research* 2:221–227
11. Lukiw WJ, Carver LA, LeBlanc HJ, Bazan NG (2000) Analysis of 1184 gene transcript levels in AD CA1 hippocampus: synaptic signaling and transcription factor deficits and upregulation of pro-inflammatory pathways. *Alzheimer's Res* 3:161–167
12. Loring JF, Wen X, Lee JM, Seilhamer J, Somogyi R (2001) Gene expression profile of Alzheimer's disease. *DNA Cell Biol* 20:683–695
13. Tsuji T, Shiozaki A, Kohno R, Yoshizato K, Shimohama S (2002) Proteomic profiling and neurodegeneration in Alzheimer's disease. *Neurochem Res* 27:1245–1253
14. Alzheimer A, Stelzmann RA, Schnitzlein HN, Murtagh FR (1995) An English translation of Alzheimer's 1907 paper, "Über eine eigenartige Erkrankung der Hirnrinde". *Clin Anat* 8:429–431
15. Butterfield DA, Drake J, Pocernich C, Castegna A (2001) Evidence of oxidative damage in Alzheimer's disease brain: central role for amyloid β -peptide. *Trends Mol Med* 7:548–554
16. Sleegers K, Van Duijn CM (2001) Alzheimer's disease: genes, pathogenesis and risk prediction. *Community Genet* 4:197–203
17. Gambalin TC, Chen F, Zambrano A, Abraha A, Lagalwar S, Guillozet AL, Lu M, Fu Y, Garcia-Sierra F, LaPointe N, Miller R, Berry RW, Binder LI, Cryns VL (2003) Caspase cleavage of tau: linking amyloid and neurofibrillary tangles in Alzheimer's disease. *Proc Natl Acad Sci* 100:10032–10037
18. Cotman CW, Poon WW, Rissman RA, Blurton-Jones M (2005) The role of caspase cleavage of tau in Alzheimer disease neuropathology. *J Neuropathol Exp Neurol* 64:104–112
19. Rohn TT, Head E (2008) Caspase activation in Alzheimer's disease: early to rise and late to bed. *Rev Neurosci* 19:383–393
20. Rohn TT (2009) Cytoplasmic inclusions of TDP-43 in neurodegenerative diseases: a potential role for caspases. *Histol Histopathol* 24:1081–1086
21. Whelan J (2008) n-6 and (n-3) polyunsaturated fatty acids and the aging brain: food for thought. *J Nutr* 138:2521–2522
22. Fukaya T, Fukaya T, Gondaira T, Kashiya Y, Kotani S, Ishikura Y, Fujikawa S, Kiso Y, Sakakibara M (2007) Arachidonic acid preserves hippocampal neuron membrane fluidity in senescent rats. *Neurobiol Aging* 28:1179–1186
23. Montecucco C, Gutiérrez JM, Lomonte B (2008) Cellular pathology induced by snake venom phospholipase A2 myotoxins and neurotoxins: common aspects of their mechanisms of action. *Cell Mol Life Sci* 65:2897–2912
24. Sun GY, Shelat PB, Jensen MB, He Y, Sun AY, Simonyi A (2009) Phospholipases A2 and inflammatory responses in the central nervous system. *Neuromolecular Med*. doi:10.1007/s12017-009-8092-z Oct 24
25. Franceschi C, Valensin S, Lescai F, Olivieri F, Licastro F, Grimaldi LM, Monti D, De Benedictis G, Bonafe M (2001) Neuroinflammation and the genetics of Alzheimer's disease: the search for a pro-inflammatory phenotype. *Aging* 13:163–170
26. Sayre LM, Smith MA, Perry G (2001) Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Curr Med Chem* 8:721–738
27. Rogers J (1995) Inflammation as a pathogenic mechanism in Alzheimer's disease. *Arzneimittelforschung* 45:439–442
28. Tol J, Roks G, Slooter AJ, van Duijn CM (1999) Genetic and environmental factors in Alzheimer's disease. *Rev Neurol* 155:10–16
29. Rogan S, Lippa CF (2002) Alzheimer's disease and other dementias: a review. *Amer J Alzheimer's Dis Other Dement* 17:11–17
30. Bazan NG, Colangelo V, Lukiw WJ (2002) Prostaglandins and other lipid mediators in Alzheimer's disease. *Prostaglandins Other Lipid Mediat* 68–69:197–210
31. Marcheselli VL, Hong S, Lukiw WJ, Tian XH, Gronert K, Musto A, Hardy M, Gimenez JM, Chiang N, Serhan CN, Bazan NG (2003) Novel docosanoids inhibit brain ischemia-reperfusion-mediated leukocyte infiltration and pro-inflammatory gene expression. *J Biol Chem* 278:43807–43817
32. Farkas E, de Wilde MC, Kiliaan AJ, Meijer J, Keijser JN, Luiten PG (2002) Dietary long chain PUFAs differentially affect hippocampal muscarinic 1 and serotonergic 1A receptors in experimental cerebral hypoperfusion. *Brain Res* 954:32–41
33. Heinemann KM, Bauer JE (2006) Docosahexaenoic acid and neurologic development in animals. *J Am Vet Med Assoc* 228:700–705
34. Lim GP, Calon F, Morihara T, Yang F, Teter B, Ubeda O, Salem N Jr, Frautschy SA, Cole GM (2005) A diet enriched with the omega-3 fatty acid docosahexaenoic acid reduces amyloid burden in an aged Alzheimer mouse model. *J Neurosci* 25:3032–3040
35. Lukiw WJ, Percy ME, Kruck TP (2005) Nanomolar metal sulfates induce pro-inflammatory and pro-apoptotic gene expression in human brain cells in primary culture. *J Inorg Biochem* 99:1895–1898
36. Lukiw WJ, Bazan NG (2006) Survival signaling in Alzheimer's disease. *Biochem Soc Trans* 34:1277–1282
37. Niemoller TD, Stark DT, Bazan NG (2009) Omega-3 fatty acid docosahexaenoic acid is the precursor of neuroprotectin D1 in the nervous system. *World Rev Nutr Diet* 99:46–54
38. Bazan NG (2009) Neuroprotectin D1-mediated anti-inflammatory and survival signaling in stroke, retinal degenerations, and Alzheimer's disease. *J Lipid Res* 50:400–405
39. Minihane AM, Khan S, Leigh-Firbank EC, Talmud P, Wright JW, Murphy MC, Griffin BA, Williams CM (2005) ApoE polymorphism and fish oil supplementation in subjects with an atherogenic lipoprotein phenotype. *Arterioscler Thromb Vasc Biol* 20:1990–1997
40. Huang TL, Zandi PP, Tucker KL, Fitzpatrick AL, Kuller LH, Fried LP, Burke GL, Carlson MC (2005) Benefits of fatty fish on dementia risk are stronger for those without APOE epsilon4. *Neurology* 65:1409–1414
41. Cole GM, Frautschy SA (2010) DHA may prevent age-related dementia. *J Nutr*. [Epub ahead of print] PubMed PMID: 20181786
42. Repici M, Mariani J, Borsello T (2007) Neuronal death and neuroprotection: a review. *Meth Mol Biol* 399:1–14
43. Gorman AM (2008) Neuronal cell death in neurodegenerative diseases: recurring themes around protein handling. *J Cell Mol Med* 12:2263–2280
44. Lorz C, Mehmet H (2009) The role of death receptors in neural injury. *Front Biosci* 14:583–595
45. DeLegge MH, Smoke A (2008) Neurodegeneration and inflammation. *Nutr Clin Pract* 23:35–41
46. Du H, Yan SS (2009) Mitochondrial permeability transition pore in Alzheimer's disease: cyclophilin D and amyloid β . *Biochim Biophys Acta* 1802:198–204
47. Kalinichenko SG, Matveeva NY (2008) Morphological characteristics of apoptosis and its significance in neurogenesis. *Neurosci Behav Physiol* 38:333–342
48. Lukiw WJ (2008) Emerging amyloid β (A β) peptide modulators for the treatment of Alzheimer's disease (AD). *Expert Opin Emerg Drugs* 13:255–271
49. Lukiw WJ, Cui JG, Marcheselli VL, Bodker M, Botkjaer A, Gotlinger K, Serhan CN, Bazan NG (2005) A role for docosahex-

- aenoic acid-derived neuroprotectin D1 in neural cell survival and Alzheimer disease. *J Clin Invest* 115:2774–2783
50. Lukiw WJ (2009) Docosahexaenoic acid and amyloid-beta peptide signaling in Alzheimer's disease. *World Rev Nutr Diet* 99:55–70
51. Bazan NG (2009) Cellular and molecular events mediated by docosahexaenoic acid-derived neuroprotectin D1 signaling in photoreceptor cell survival and brain protection. *Prostaglandins Leukot Essent Fatty Acids* 81:205–211
52. Lukiw WJ, Bazan NG (2008) Docosahexaenoic acid and the aging brain. *J Nutr* 138:2510–2514
53. Zhao Y, Cui JG, Lukiw WJ (2006) Natural secretory products of human neural and microvessel endothelial cells: implications in pathogenic “spreading” and Alzheimer's disease. *Mol Neurobiol* 34:181–192
54. McGeer PL, Rogers J, McGeer EG (2006) Inflammation, anti-inflammatory agents and Alzheimer disease: the last 12 years. *J Alzheimers Dis* 9:271–276